



# OptiBst DNA Polymerase

(Large fragment, exo-)  
(*Bacillus stearothermophilus*)

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Cat. No.	Size
E1079-01	1000 units
E1079-02	8000 units

**Unit Definition:** One unit is the amount of enzyme required to incorporate 10 nmoles of total deoxyribonucleotide into acid-insoluble form in 30 min at 60°C.

**Storage Conditions:**  
Store at -20°C

## Thermostable *Bst* DNA Polymerase (large fragment, exo-), exhibiting strand displacement activity.

### Description:

- *Bst* DNA Polymerase is a moderately thermostable enzyme from *Bacillus stearothermophilus*.
- The enzyme is genetically optimized for faster amplification and more flexibility towards reaction conditions.
- Optimized for LAMP (Loop Mediated Isothermal Amplification).
- Enhanced capability to incorporate dUTP, as compared to wild-type *Bst* DNA Polymerase
- Ultrapure, recombinant protein.
- The enzyme replicates DNA optimally at 65°C.
- Catalyzes the polymerization of nucleotides into duplex DNA in the 5'→3' direction in the presence of magnesium ions.
- Lacks the 5'→3' exonuclease activity, while retaining polymerase activity (1).
- Maintains DNA strand displacement activity.
- Broad activity range. Can replace mesophilic polymerases as well as synthesize DNA at elevated temperatures. Thus it is suitable for amplification of difficult DNA templates, including repetitive sequences, GC-rich regions and problematic secondary structures (2, 3).
- Heat inactivation at temperatures above 80°C.
- Active over a wide range of reaction buffer conditions and magnesium ions concentrations.
- Used in isothermal DNA sequencing at elevated temperatures.
- Ideal for DNA synthesis reactions requiring strand displacement

### Storage Buffer:

10 mM Tris-HCl (pH 8.0 at 20°C), 50 mM KCl, 0.1 mM EDTA, 0.1% Tergitol TMN, 1 mM DTT and 50% (v/v) glycerol.

### 1x Opti Bst DNA Polymerase Reaction Buffer:

50 mM Tris-HCl (pH 8.9 at 20°C), 10 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 50 mM KCl, 2 mM-MgSO<sub>4</sub>, 0.1% Tergitol TMN.

### Quality Control:

All preparations are assayed for contaminating endonuclease, exonuclease, and single- and double-stranded DNase activities.

### References:

- (1) Stenesh, J. and Roe, B.A. (1972) *Biochim. Biophys. Acta.* 272, 156-166.
- (2) Hugh, G. and Griffin, M. (1994) *PCR Technology*, p.p.228-229.
- (3) McClary, J. et al. (1991) *J. DNA Sequencing and Mapping*, p.p.173-180